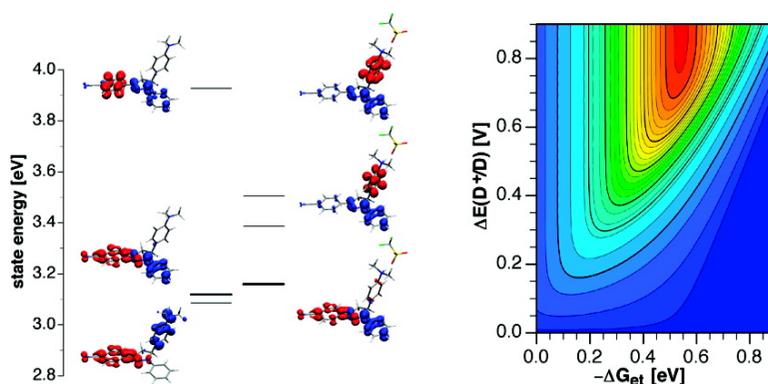


Differential Tuning of the Electron Transfer Parameters in 1,3,5-Triarylpyrazolines: A Rational Design Approach for Optimizing the Contrast Ratio of Fluorescent Probes

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Differential Tuning of the Electron Transfer Parameters in 1,3,5-Triarylpyrazolines: A Rational Design Approach for Optimizing the Contrast Ratio of Fluorescent Probes

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Abstract: A large class of cation-responsive fluorescent sensors utilizes a donor–spacer–acceptor (D–A) molecular framework that can modulate the fluorescence emission intensity through a fast photoinduced intramolecular electron transfer (PET) process. The emission enhancement upon binding of the analyte defines the contrast ratio of the probe, a key property that is particularly relevant in fluorescence microscopy imaging applications. Due to their unusual electronic structure, 1,3,5-triarylpyrazoline fluorophores allow for the differential tuning of the excited-state energy ΔE_{00} and the fluorophore acceptor potential $E(A/A^-)$, both of which are critical parameters that define the electron transfer (ET) thermodynamics and thus the contrast ratio. By systematically varying the number and attachment positions of fluoro substituents on the fluorophore π -system, ΔE_{00} can be adjusted over a broad range (0.4 eV) without significantly altering the acceptor potential $E(A/A^-)$. Experimentally measured D–A coupling and reorganization energies were used to draw a potential map for identifying the optimal ET driving force that is expected to give a maximum fluorescence enhancement for a given change in donor potential upon binding of the analyte. The rational design strategy was tested by optimizing the fluorescence response of a pH-sensitive probe, thus yielding a maximum emission enhancement factor of 400 upon acidification. Furthermore, quantum chemical calculations were used to reproduce the experimental trends of reduction potentials, excited-state energies, and ET driving forces within the framework of linear free energy relationships (LFERs). Such LFERs should be suitable to semiempirically predict ET driving forces with an average unsigned error of 0.03 eV, consequently allowing for the computational prescreening of substituent combinations to best match the donor potential of a given cation receptor. Within the scaffold of the triarylpyrazoline platform, the outlined differential tuning of the electron transfer parameters should be applicable to a broad range of cation receptors for designing PET sensors with maximized contrast ratios.

Introduction

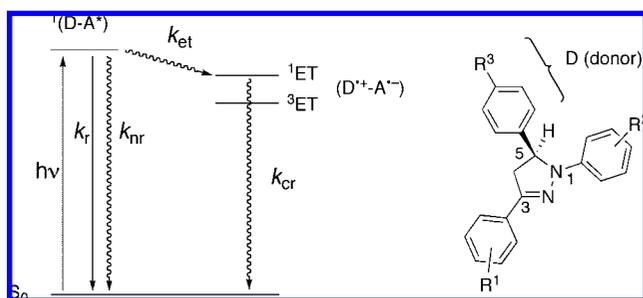
Fluorescent probes have found widespread applications in cell biology for visualizing the dynamics of intracellular processes.¹ An increasing body of research has been devoted to the development of fluorescent probes for the noninvasive measurement of biologically relevant metal cations, such as calcium, magnesium, zinc, or copper, as well as the detection of hazardous heavy metals, including mercury, lead, and cadmium.² A large class of these sensors utilizes a donor–spacer–acceptor (D–A) molecular framework that can modulate the fluorescence emission intensity through a fast photoinduced intramolecular electron transfer (PET) process.³ In the absence of the cation, fluorescence emission is quenched through PET from the donor to the fluorophore acting as an electron acceptor. Upon binding of the cation to the donor moiety, the PET process is rendered energetically less favorably, resulting in a fluorescence increase.

Although the photophysical mechanism that dictates the “switch on” behavior of PET sensors is well understood,⁴ many successfully developed cation sensors have been made on the basis of empirical principles rather than rational design. More

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Scheme 1



recently, tuning of the frontier orbital energy levels involved in the photoinduced electron transfer process has been used as a key concept for the design of a broad range of PET sensors.^{5–7} The optimization of the fluorescence enhancement upon binding of the analyte is particularly important, because it defines the optical sensitivity and contrast ratio in microscopy imaging applications. The goal of this study was to delineate a generalized rational design approach for finely tuning the fluorophore properties and thus optimizing the contrast ratio for a given cation receptor.

A number of photophysical parameters govern the fluorescence enhancement of a PET sensor upon analyte binding. The fluorescence quantum yield Φ_f^0 of the fluorophore in the absence of a quenching electron donor is determined by the radiative and nonradiative deactivation rate constants k_r and k_{nr} , respectively:⁸

$$\Phi_f^0 = \frac{k_r}{k_r + k_{nr}} \quad (1)$$

In the presence of an electron donor (D), an additional nonradiative deactivation pathway is introduced that competes with excited-state deactivation as illustrated with the simplified Jablonski diagram in Scheme 1 (left). The singlet excited state $^1(D-A^*)$ can undergo a fast intramolecular electron transfer reaction in which the fluorophore is acting as an electron acceptor (A), yielding a radical ion pair ($D^{+}A^{-}$). Charge recombination is in most cases a nonradiative process, regenerating the original ground-state species. Depending on the rate k_{et} of electron transfer (ET), fluorescence emission is quenched and the quantum yield is lowered from Φ_f^0 to Φ_f :

$$\Phi_f = \frac{k_r}{k_r + k_{nr} + k_{et}} = \frac{\Phi_f^0 k_0}{k_0 + k_{et}} \quad \text{with} \quad k_0 = k_r + k_{nr} \quad (2)$$

If a rigid σ -bond spacer is used to connect the electron donor with the fluorophore π -system, electronic coupling between the two units is expected to be small.⁹ In this case, the electron transfer kinetics is best described as a reaction in the nonadiabatic regime using semiclassical Marcus theory.¹⁰ Within this framework, the electron transfer rate constant k_{et} depends on the thermodynamic driving force $-\Delta G_{et}$ of the ET reaction, the associated reorganization energy λ , and the electronic coupling

H_{DA} between the excited precursor state ($^1(D-A^*)$) and the successor radical ion pair $D^{+}A^{-}$:

$$k_{et} = \left(\frac{4\pi^3}{h^2 \lambda k_B T} \right)^{1/2} H_{DA}^2 \exp \left[-\frac{(\Delta G_{et} + \lambda)^2}{4\lambda k_B T} \right] \quad (3)$$

with k_B = Boltzman constant, T = temperature, and h = Planck constant.

The driving force of the ET process ($-\Delta G_{et}$) in turn can be experimentally estimated by the Rehm–Weller equation (4),¹¹ which relates the free energy change ΔG_{et} with the donor and acceptor ground-state potentials $E(D^+/D)$ and $E(A/A^-)$, respectively, the zero-zero transition energy ΔE_{00} of the fluorophore, and a work term, w_p , corresponding to the Coulomb stabilization energy of the formed radical ion pair in the respective solvent.

$$\Delta G_{et} = E(D^+/D) - E(A/A^-) - \Delta E_{00} + w_p \quad (4)$$

Coordination of a cation to the electron-donating moiety of the PET sensor results in an increase of the donor potential $E(D^+/D)$ and reorganization energy λ . As a consequence, the ET driving force $-\Delta G_{et}$ is lowered (eq 4), the electron transfer rate reduced (eq 3), and the fluorescence quantum yield increased (eq 2). Denoting the new quantum yield and ET rate constant as Φ_f' and k_{et}' , respectively, the observed fluorescence enhancement f_e or contrast ratio can be expressed as

$$f_e = \frac{\Phi_f'}{\Phi_f} = \frac{k_0 + k_{et}}{k_0 + k_{et}'} \quad (5)$$

Hence, to maximize f_e , the difference of the ET rates in the presence and absence of the analyte must be optimized. This can be principally accomplished by adjusting the driving force $-\Delta G_{et}$ through changes in $E(A/A^-)$ and ΔE_{00} of the fluorophore or $E(D^+/D)$ of the cation receptor acting as the electron donor. Because the cation binding site is electronically decoupled from the fluorophore π -system, the latter parameter can be varied without significantly affecting $E(A/A^-)$ and ΔE_{00} . In many cases, however, the cation binding site has been purposefully designed to achieve high selectivity toward a particular analyte, and therefore, adjusting the donor potential $E(D^+/D)$ without changing the binding affinity and selectivity of the receptor site is a nontrivial task. For this reason, tuning of either the acceptor potential $E(A/A^-)$ or ΔE_{00} would be more desirable. However, this approach is equally challenging, because both parameters are defined by the fluorophore structure and attempts to tune a single parameter are expected to inevitably affect the other. Recent studies demonstrated that these two parameters can be independently adjusted in 1,3,5-triarylpyrazoline-based fluorophores.⁷ As shown in Scheme 1 (right), two of the aryl rings in this fluorophore communicate electronically through the pyrazoline π -system, whereas the third ring in the 5-position is electronically decoupled and can be utilized as an electron-donating cation receptor.^{6,12} Substituents attached to the phenyl ring in the 1-position have a strong effect on the excited-state energy but only a small influence on the reduction potential of

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the fluorophore. This observation prompted us to investigate whether 1,3,5-triarylpyrazolines could be utilized for optimizing the contrast ratio of PET sensors in a systematic and predictable manner.

Experimental Section

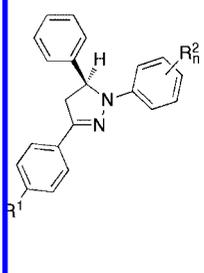
Synthesis. All compounds were prepared following protocols from the literature.^{7,13} The crude products were purified by flash chromatography, and the purities of the final products were confirmed by reversed-phase HPLC (Varian ProStar system with UV detector, acetonitrile–water, gradient 20% → 2% water). The chemical structures of the synthesized compounds were confirmed by ¹H NMR, ¹³C NMR, MS, and high resolution mass spectrometry. A description of the syntheses and detailed analytical data are provided in the Supporting Information.

Absorption and Fluorescence Spectroscopy. All sample solutions were filtered through 0.45 μm Teflon membrane filters to remove interfering dust particles or fibers. UV–vis absorption spectra were recorded at 25 °C using a Varian Cary Bio50 UV–vis spectrometer with constant-temperature accessory. Steady-state emission and excitation spectra were recorded with a PTI fluorimeter. For all measurements the path length was 1 cm with a cell volume of 3.0 mL. The fluorescence spectra have been corrected for the spectral response of the detection system (emission correction file provided by the instrument manufacturer) and for the spectral irradiance of the excitation channel (via a calibrated photodiode). Quantum yields were determined using quinine sulfate dihydrate in 1.0 N H₂SO₄ as a fluorescence standard ($\Phi_F = 0.54 \pm 0.05$).¹⁴ Time-resolved fluorescence decay data were acquired with a single photon counting spectrometer (Edinburgh Instruments, LifeSpec Series, pulsed laser diode excitation at 370 nm, fwhm = 80 ps).

Cyclic Voltammetry. The cyclic voltammograms were acquired in acetonitrile (freshly distilled from calcium hydride) containing 0.1 M Bu₄NPF₆ as an electrolyte using a CH-Instruments potentiostat (model 600A). The samples were measured under an inert gas at a concentration of 3 mM in a single-compartment cell with a glassy carbon working electrode, a Pt counter electrode, and a Ag/AgNO₃ (10 mM in CH₃CN) nonaqueous reference electrode. The half-wave potentials were referenced to ferrocene as an internal standard. All measurements were performed with a scan rate of 100 mV s⁻¹.

Computational Methods. All calculations were carried out with the Q-Chem electronic structure calculation suite of programs.¹⁵ Ground-state geometries were energy minimized by DFT with the B3LYP hybrid functional¹⁶ and Pople's 6-31+G(d) split valence basis set with added diffuse and polarization functions. The starting geometry for each optimization was constructed from the X-ray coordinates of unsubstituted 1,3,5-triphenylpyrazoline.¹⁷ To ensure a stationary point on the ground-state potential surface, all geometry-optimized structures were verified by a vibrational frequency analysis, which at the same time also provided zero-point vibrational energy (ZPVE) corrections. Adiabatic gas-phase electron affinities (EAs) were calculated as the difference between the total energies of the geometry-optimized anionic and charge-neutral fluorophores. The electronic energies were corrected with the respective scaled harmonic zero-point vibrational energies (scaling factor 0.9806).¹⁸ To obtain estimates of the vertical electronic excitation energies which include some account of electron correlation, time-dependent density functional theory (TD-DFT)

Chart 1



	R ¹ = H	R ¹ = CN	R _n ²
1a	2a	H	
1b	2b	2-F	
1c	2c	3-F	
1d	2d	4-F	
1e	2e	2,4-F ₂	
1f	2f	2,5-F ₂	
1g	2g	3,5-F ₂	
1h	2h	2,4,5-F ₃	
1i	2i	2,3,5,6-F ₄	
1j	2j	2,3,4,5,6-F ₅	

calculations¹⁹ with the B3LYP functional and the 6-31+G(d) or 6-311+G(2d,2p) basis set were performed. Molecular orbitals were visualized with the software VMD²⁰ using the Q-Chem plot output data. Details of the computational results including the coordinates of the geometry-optimized structures are provided in the Supporting Information.

Results and Discussion

To systematically explore the tunability of the photophysical properties of 1,3,5-triarylpyrazolines in the absence of a quenching electron donor (D), we first synthesized a small compound library composed of derivatives with increasing number of fluorine substituents attached to various positions of the *N*-aryl ring (Chart 1). The effect of substituents on the 3-aryl ring was probed by comparing the parent compound series **1a–j** with the cyano-substituted compounds **2a–j**. All derivatives were synthesized in two steps as previously published.⁷ Aldol condensation of benzaldehyde with the acetyl derivative yielded a chalcone intermediate, which was converted to the desired racemic pyrazolines by cyclization with the corresponding fluoro-substituted phenylhydrazine derivative.

Steady-State Absorption and Emission Spectroscopy. A compilation of the photophysical properties for compounds **1a–j** and **2a–j** is given in Table 1. All data were measured at 298 K in acetonitrile as the solvent. Within the applied concentration range, normalized absorption spectra were superimposable and scaled linearly, indicating no detectable aggregation. Collectively, the two series cover a broad range in absorption ($\Delta\lambda_{\max} = 82$ nm) and emission ($\Delta\lambda_{\max} = 105$ nm) maxima (Figure 1). With increasing number of fluoro substituents, the absorption and emission maxima were shifted to higher energy. This trend correlates well with the anticipated degree of electron withdrawing character imposed by the fluoro substituents, which gradually reduce charge delocalization from the 1-*N*-pyrazoline nitrogen lone pair toward the 3-aryl ring in the ground and excited states.⁷ The zero–zero transition energy ΔE_{00} , one of the key parameters for tuning the ET thermodynamics (eq 4), varies over nearly 6000 cm⁻¹ or 0.73 eV. Within each compound series **1** and **2**, the tunable range covers more than 3000 cm⁻¹ or 0.4 eV. A comparison of the Stokes shift revealed only small variations for both series with an average of approximately 6100 ± 500 cm⁻¹, thus indicating similar degrees of excited-state geometrical relaxation and solvent stabilization. The rather uniform half-width of the normalized absorption and emission spectra further supports similar shapes for the ground- and excited-state

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Table 1. Photophysical Data of Pyrazoline Derivatives **1a–j** and **2a–j** in Acetonitrile at 298 K

	abs λ_{\max} (nm)	em λ_{\max} (nm)	Stokes shift (cm ⁻¹)	ΔE_{00} ^a (cm ⁻¹)	Φ_f ^b		abs λ_{\max} (nm)	em λ_{\max} (nm)	Stokes shift (cm ⁻¹)	ΔE_{00} (cm ⁻¹)	Φ_f ^b
1a	356	459	6300	24 940	0.65	2a	396	508	5560	22 470	0.30
1b	344	445	6580	25 770	0.64	2b	378	496	6280	23 310	0.58
1c	352	440	5700	25 560	0.65	2c	388	492	5430	23 060	0.63
1d	357	457	6110	24 960	0.60	2d	393	512	5890	22 500	0.50
1e	342	447	6880	25 800	0.55	2e	374	496	6580	23 450	0.59
1f	345	434	5930	26 020	0.68	2f	376	485	5970	23 600	0.63
1g	347	427	5410	26 120	0.70	2g	381	476	5240	23 630	0.68
1h	344	437	6220	25 960	0.64	2h	371	488	6460	23 720	0.62
1i	324	401	5910	27 920	0.04	2i	351	447	6110	25 420	0.79
1j	314	403	7070	28 320	<0.01	2j	343	449	6900	25 700	0.71

^a Zero-zero transition energy, estimated on the basis of $\Delta E_{00} = ((E_{\text{abs}}(\text{max}) + E_{\text{em}}(\text{max}))/2)$. ^b Fluorescence quantum yield, quinine sulfate as a reference.

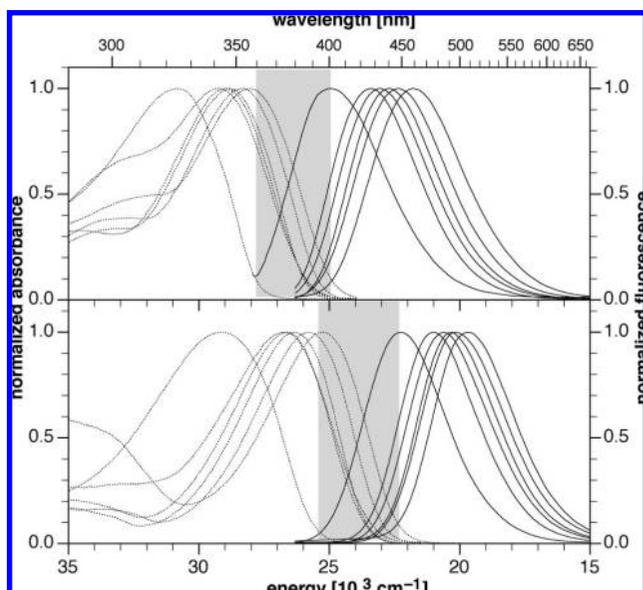


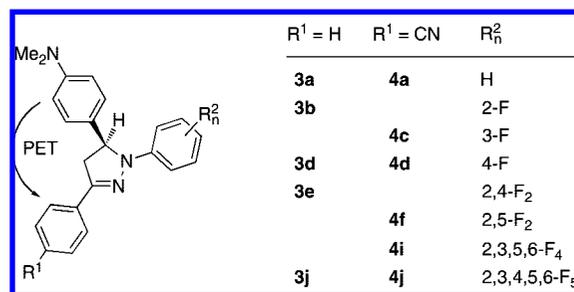
Figure 1. Comparison of the normalized absorption and emission spectra of compounds **1a–j** (top) and **2a–j** (bottom) in acetonitrile. Absorption spectra are shown as dotted lines, and emission spectra are shown as solid lines. The shaded areas indicate the tunable range of the excited-state energy ΔE_{00} for each compound series.

potential energy surfaces across all fluorophores (Figure 1). With the exception of **1i** and **1j**, all compounds fluoresce brightly with an average quantum yield Φ_f^0 of 0.6 ± 0.1 .

Electrochemistry. To evaluate changes in the acceptor potential $E(\text{A}/\text{A}^-)$ of the fluorophore, the second key parameter governing the ET thermodynamics according to eq 4, we determined the half-wave potentials for each compound by cyclic voltammetry in acetonitrile and 0.1 M Bu_4NPF_6 as the electrolyte. As evident from the compilation in Table 2, the acceptor potentials differed significantly between the two series **1** and **2** but showed surprisingly little variation within each series. More specifically, for compounds **1a–j**, the average reduction potential was centered around -2.74 ± 0.05 V, while for **2a–j**, which contain the strongly electron withdrawing cyano group, a less negative potential of -2.21 ± 0.03 V was measured. In both series, the fluoro substituents exerted only a small influence on the reduction potential, an observation that is rooted in the special topology of the HOMO and LUMO in this class of fluorophores.⁷ In contrast, the excited-state energy ΔE_{00} is strongly influenced by the number of fluoro substituents, making it possible to tune ΔE_{00} over 0.4 eV while locking in the reduction potential within a narrow range. It is noteworthy that this property allows for the design of two fluorophores with

Table 2. Acceptor Half-Wave Reduction Potentials for Pyrazoline Derivatives **1a–j** and **2a–j** in Acetonitrile/0.1 M Bu_4NPF_6 vs $\text{Fc}^{+/0}$ at 298 K

compd	$E_{1/2}(\text{A}/\text{A}^-)$ (V)	compd	$E_{1/2}(\text{A}/\text{A}^-)$ (V)
1a	-2.79	2a	-2.26
1b	-2.78	2b	-2.20
1c	-2.76	2c	-2.21
1d	-2.77	2d	-2.24
1e	-2.75	2e	-2.21
1f	-2.70	2f	-2.18
1g	-2.71	2g	-2.16
1h	-2.69	2h	-2.17
1i	-2.72	2i	-2.21
1j	-2.66	2j	-2.22

Chart 2

essentially identical excited-state energies but with reduction potentials that differ by more than 0.5 V as illustrated with **1b** and **2j**. Hence, by varying the number and positions of fluoro substituents, the excited-state energy ΔE_{00} can be gradually adjusted over a broad range without significantly altering the acceptor potential $E(\text{A}/\text{A}^-)$, a remarkable characteristic that might be exploited for the systematic tuning of the ET driving force $-\Delta G_{\text{et}}$ according to eq 4.

Optimizing the Fluorescence Enhancement in PET Sensors.

As outlined above, tuning of the ET driving force $-\Delta G_{\text{et}}$ is pivotal for optimizing the contrast ratio of PET sensors. To explore whether the differential tuning strategy can be used for the latter purpose, we next synthesized a compound series containing a dimethylamino group which serves as the quenching electron donor (Chart 2).²¹ Protonation of the amino nitrogen lone pair is expected to sharply increase the donor potential, which in turn should result in a quantum yield increase. Thus, the dimethylamino-substituted compound series serves as a simple pH-responsive PET model system to test the rational design approach. The compound numbering scheme of this

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Table 3. Photophysical Data of Pyrazoline Derivatives **3** and **4** in Acetonitrile at 298 K

	abs λ_{max} (nm)	em λ_{max} (nm)	Stokes shift (cm^{-1})	ΔE_{00}^a (cm^{-1})	Φ_f^b	$\Phi_f'^c$	f_e^d
3a	357	455	6030	24 990	0.44	0.29	0.66
3b	344	448	6750	25 700	0.37	0.25	0.68
3d	358	458	6100	24 880	0.45	0.22	0.49
3e	343	450	6930	25 690	0.29	<0.01	nd
4a	400	508	5320	22 340	0.014	0.49	35
4c	392	488	5020	23 000	0.0076	0.61	80
4d	396	512	5720	22 390	0.016	0.51	33
4f	379	481	5600	23 590	0.0038	0.63	166
4i	356	445	5620	25 280	0.0033	0.71	215
4j	349	446	6230	25 540	0.0017	0.68	400

^a Zero-zero transition energy, estimated on the basis of $\Delta E_{00} = ((E_{\text{abs}}(\text{max}) + E_{\text{em}}(\text{max}))/2)$. ^b Fluorescence quantum yield in acetonitrile. ^c Fluorescence quantum yield in 10 mM trifluoroacetic acid in acetonitrile, quinine sulfate as a reference. ^d Fluorescence enhancement upon acidification $f_e = \Phi_f'/\Phi_f$.

series was kept consistent with the first library (Chart 1), although a significantly smaller number of fluoro derivatives were included.

As evident from the compilation in Table 3, compared to the unsubstituted parent compounds of series **1** and **2**, the absorption and emission maxima are not significantly altered by attachment of the dimethylamino group. However, a comparison of the quantum yields in neutral solution revealed significant changes. Notably, the fluorescence of the cyano-substituted compounds **4a–j** is almost entirely quenched in the presence of the dimethylamino group. Upon acidification with 10 mM trifluoroacetic acid in acetonitrile, the quantum yields Φ_f' sharply increased for compounds **4a–j**, yielding fluorescence enhancement factors of up to 400. Interestingly, the quantum yields of the 3-phenyl-substituted derivatives **3a–e** were consistently lower in acidic compared to neutral medium. This unexpected observation might be a result of the strongly electron withdrawing character of the protonated aniline, which may act as an electron acceptor in its protonated form (vide infra).

Electron Transfer Driving Force and Kinetics. Assuming that the reduced quantum yield Φ_f of the dimethylamino-substituted fluorophores relative to their unsubstituted parent compounds (Φ_f^0) is entirely a consequence of the ET reaction, the ET rate constant k_{et} can be estimated on the basis of eq 2. This approximation implies that the kinetics of all other decay pathways (radiative deactivation, internal conversion, intersystem crossing) remains unchanged upon substitution with the donor group. Solving eq 2 for k_{et} then gives

$$k_{\text{et}} = \frac{1}{\tau_f} \left(\frac{\Phi_f^0}{\Phi_f} - 1 \right) \quad \text{with} \quad \tau_f = \frac{1}{k_0} \quad (6)$$

The excited-state deactivation rate constant k_0 can be directly obtained from the fluorescence lifetime data τ_f of the parent compound. Additionally, to estimate the electron transfer driving force $-\Delta G_{\text{et}}$ according to eq 4, the donor and acceptor half-wave potentials $\Delta E(\text{D}^+/\text{D})$ and $\Delta E(\text{A}/\text{A}^-)$ as well as the excited-state energy ΔE_{00} for each compound of series **3** and **4** were experimentally determined in acetonitrile as the solvent. The Coulomb stabilization energy of the radical ion pair was estimated according to the following equation:

$$w_p = -\frac{e^2}{\epsilon_s d_{\text{DA}}} \quad (7)$$

where e stands for the elemental charge, ϵ_s for the static dielectric constant, and d_{DA} for the separation between the ion

Table 4. Donor and Acceptor Reduction Potentials and Electron Transfer Parameters of Compounds **3** and **4** in Acetonitrile at 298 K

	$E_{1/2}(\text{D}^+/\text{D})^a$ (V)	$E_{1/2}(\text{A}/\text{A}^-)^a$ (V)	τ_f^b (ns)	k_0^c (10^8 s^{-1})	E_{00}^d (eV)	ΔG_{et}^e (eV)	k_{et}^f (s^{-1})
3a	0.38	-2.89	4.29	2.33	3.09	0.13	1.1×10^8
3b	0.42	-2.86	4.24	2.36	3.19	0.05	1.7×10^8
3d	0.38	-2.87	4.52	2.21	3.09	0.12	7.4×10^7
3e	0.43	-2.82	4.43	2.26	3.20	0.02	2.0×10^8
4a	0.43	-2.26	3.82	2.62	2.79	-0.13	5.3×10^9
4c	0.44	-2.23	3.98	2.51	2.86	-0.23	2.0×10^{10}
4d	0.44	-2.26	3.97	2.52	2.79	-0.12	7.8×10^9
4f	0.45	-2.21	3.99	2.51	2.93	-0.31	4.1×10^{10}
4i	0.49	-2.23	3.09	3.24	3.15	-0.46	7.7×10^{10}
4j	0.47	-2.18	3.03	3.30	3.19	-0.56	1.4×10^{11}

^a Half-wave potential in acetonitrile/0.1 M Bu₄NPF₆ vs Fe^{+/0} at 298 K. ^b Fluorescence lifetime of the corresponding reference compounds **2** and **3** having the same fluoro substitution pattern but lacking the dimethylamino donor group. All decay profiles fitted well with a monoexponential kinetics ($\chi^2 < 1.3$). ^c Excited-state deactivation rate constant $k_0 = 1/\tau_f$. ^d Zero-zero transition energy, estimated on the basis of $\Delta E_{00} = ((E_{\text{abs}}(\text{max}) + E_{\text{em}}(\text{max}))/2)$. ^e Calculated on the basis of the Rehm–Weller equation (4) with $w_p = -0.045 \text{ eV}$. ^f Electron transfer rate constant calculated on the basis of eq 6.

pair. In a polar environment such as acetonitrile, this contribution is relatively small compared to the other parameters. Using an average D–A separation of 8.6 Å for the two compound series **3** and **4**, a Coulomb stabilization energy of -0.045 eV was included in estimating ΔG_{et} . Because all photophysical and electrochemical data were acquired in the same solvent, the electrochemical potentials were used without additional solvent stabilization energy corrections. A compilation of all relevant data is provided in Table 4.

Given the large difference in reduction potentials, $-\Delta G_{\text{et}}$ is considerably more favorable for the cyano-substituted compound series **4** compared to series **3**. Furthermore, within a series, $-\Delta G_{\text{et}}$ steadily increases with increasing number of fluoro substituents, thus confirming the ability to tune $-\Delta G_{\text{et}}$ in a predictable manner by altering the number of fluoro substituents. The observed trend in $-\Delta G_{\text{et}}$ directly parallels the electron withdrawing character of the *N*-aryl ring. For example, due to effective resonance delocalization, a fluoro substituent in the para position as in compound **4d** is less electron withdrawing compared to one in the meta position as in compound **4c**, rendering $-\Delta G_{\text{et}}$ for **4c** more favorable compared to that for **4d**. The donor potentials are centered at $0.43 \pm 0.03 \text{ V}$ and vary only to a small degree across both compound series, an observation that is consistent with preferential oxidation of the electronically decoupled aniline moiety rather than the fluorophore π -system. Additionally, time-resolved fluorescence data in acetonitrile revealed a monoexponential decay kinetics for all reference compounds **2** and **3** with lifetimes ranging between 3 and 4.5 ns (Table 4).

A plot of $\log k_{\text{et}}$ vs $-\Delta G_{\text{et}}$ from Table 4 reveals a parabolic relationship with a more than 3 orders of magnitude increase in the ET rate as a function of the driving force (Figure 2). Nonlinear least-squares fitting of the experimental data with the semiclassical Marcus equation (3) provided the reorganization energy $\lambda = 0.54 \pm 0.04 \text{ eV}$ and the electronic coupling element $H_{\text{DA}} = 18 \pm 3 \text{ cm}^{-1}$ for the triaryl-substituted pyrazolines. The small values for the reorganization energy and electronic coupling are expected for a D–A system with a rigid spacer and compare well with the ET parameters recently estimated for fluorescein.⁹

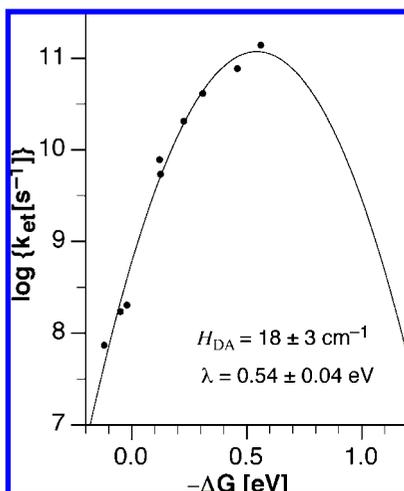


Figure 2. Dependence of the electron transfer rate constant on the driving force $-\Delta G_{\text{et}}$ in dimethylamino-substituted pyrazolines **3** and **4**. The parabolic curve was obtained through nonlinear least-squares fitting of the data according to the Marcus equation (3).

Figure 2 indicates that all ET quenching rate constants are located within the normal region of the Marcus parabola for which the driving force $-\Delta G_{\text{et}}$ is smaller than the reorganization energy λ . The driving force $-\Delta G_{\text{et}}$ is greatest for **4j**, which contains a fully fluorinated 1-aryl ring. Because none of the compounds offer a driving force beyond the parabola peak, it remains open whether the Marcus inverted region could be accessed within the framework of donor-substituted triarylpyrazolines. Due to the lack of kinetic data with larger driving forces, the ET parameters derived from the above curve fit should be considered as estimates with potentially larger uncertainties than implied by the mathematical fit. Nevertheless, the extracted parameters still offer an opportunity to explore some of the boundary conditions for this system. On the basis of the estimated reorganization energy and electronic coupling matrix element, the maximum ET rate constant k_{et} can be calculated as

$$k_{\text{et}}^{\text{max}} = \left(\frac{4\pi^3}{h^2 \lambda k_{\text{B}} T} \right)^{1/2} H_{\text{DA}}^2 = 1.2 \times 10^{11} \text{ s}^{-1} \quad (8)$$

The maximum fluorescence enhancement is reached when the ET rate of the protonated form is insignificant compared to the deactivation rate k_0 . Thus, substituting eq 5 with $k_0 + k_{\text{et}}' \approx k_0$, the maximum fluorescence enhancement can be estimated by

$$f_{\text{c}}^{\text{max}} = \frac{k_0 + k_{\text{et}}^{\text{max}}}{k_0} \quad (9)$$

Assuming an average deactivation rate constant of $k_0 = 2.8 \times 10^8 \text{ s}^{-1}$ and $k_{\text{et}} = 1.2 \times 10^{11} \text{ s}^{-1}$, the maximum fluorescence enhancement for this class of pyrazoline derivatives is expected to reside around 400.

Despite the fact that the increase in donor potential $E(\text{D}^+/\text{D})$ upon protonation of the dimethylamino group is expected to be similar for all compounds, the fluorescence enhancement varies widely within the two compound series and strongly depends on the ET driving force. For smaller $-\Delta G_{\text{et}}$ values, the quenching efficiency is lower, which results in an increased quantum yield of the PET sensor and therefore a smaller fluorescence enhancement under neutral conditions. The ex-

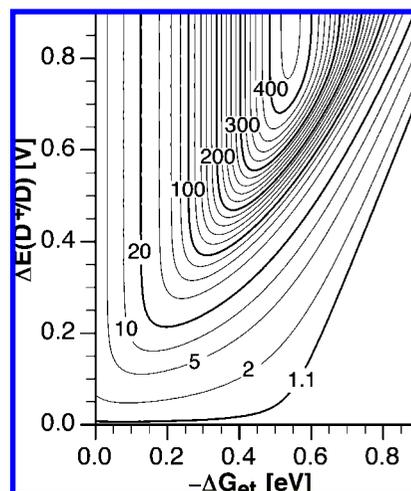


Figure 3. Contour plot of the fluorescence enhancement factor f_{c} as a function of the ET driving force and change in donor potential $\Delta E(\text{D}^+/\text{D})$ calculated according to eq 11. For f_{c} between 20 and 420 contour lines are drawn in intervals of 20. Variables: $k_0 = 2.8 \times 10^8 \text{ s}^{-1}$, $H_{\text{DA}} = 18 \text{ cm}^{-1}$, $\lambda = 0.54 \text{ eV}$, $T = 298 \text{ K}$.

perimental data imply that for a given change in donor potential $\Delta E(\text{D}^+/\text{D})$ upon binding of the analyte, in this case a simple protonation, the optimal fluorescence enhancement requires a matching ET driving force $-\Delta G_{\text{et}}$. Assuming that the change in the ET rate is predominantly caused by an increase in the donor potential ($\Delta E(\text{D}^+/\text{D})$), thus neglecting differences in the reorganization energy λ and electronic coupling H_{DA} , the relationship between $\Delta E(\text{D}^+/\text{D})$ and the ET driving force $-\Delta G_{\text{et}}$ in the absence of an analyte can be expressed through eqs 3 and 5, yielding

$$f_{\text{c}} = f(\Delta G_{\text{et}}, \Delta E(\text{D}^+/\text{D})) \\ = \frac{k_0 + \left(\frac{4\pi^3}{h^2 \lambda k_{\text{B}} T} \right)^{1/2} H_{\text{DA}}^2 \exp \left[-\frac{(\Delta G_{\text{et}} + \lambda)^2}{4\lambda k_{\text{B}} T} \right]}{k_0 + \left(\frac{4\pi^3}{h^2 \lambda k_{\text{B}} T} \right)^{1/2} H_{\text{DA}}^2 \exp \left[-\frac{(\Delta G_{\text{et}} + \Delta E(\text{D}^+/\text{D}) + \lambda)^2}{4\lambda k_{\text{B}} T} \right]} \quad (10)$$

Rewriting eq 10 allows for the expression of $\Delta E(\text{D}^+/\text{D})$ as a function of the fluorescence enhancement f_{c} and the ET driving force $-\Delta G_{\text{et}}$ of the analyte-free PET sensor:

$$\Delta E(\text{D}^+/\text{D}) = f(f_{\text{c}}, \Delta G_{\text{et}}) \\ = \left(-4\lambda k_{\text{B}} T \ln \left[\frac{(1-f_{\text{c}})}{f_{\text{c}}} \frac{k_0}{H_{\text{DA}}^2} \left(\frac{h^2 \lambda k_{\text{B}} T}{4\pi^3} \right)^{1/2} + \frac{1}{f_{\text{c}}} \exp \left[-\frac{(\Delta G_{\text{et}} + \lambda)^2}{4\lambda k_{\text{B}} T} \right] \right] \right)^{1/2} - \lambda - \Delta G_{\text{et}} \quad (11)$$

Thus, eq 11 shows the specific change in donor potential $\Delta E(\text{D}^+/\text{D})$ required to achieve a certain fluorescence enhancement f_{c} on the basis of the ET driving force $-\Delta G_{\text{et}}$ of the quenched sensor in the absence of an analyte. Conversely, if $\Delta E(\text{D}^+/\text{D})$ is known, the relationship indicates what ET driving force would offer an optimal fluorescence enhancement. A contour plot of the function in eq 11 for various enhancement factors f_{c} using the previously estimated experimental ET parameters graphically illustrates this relationship (Figure 3). Because potentially significant changes in the solvent reorga-

nization energy were not taken into account, the following interpretations should be viewed as estimates within the simplified framework of eq 11. With small potential changes (<0.1 V), a relatively broad range of ET driving forces leads already to a 2–5-fold fluorescence enhancement. Larger fluorescence enhancements not only require a greater change in donor potential, but gradually decrease the range of tolerated ET driving forces. For practical purposes, a fluorescence enhancement factor of at least 50 is desirable. To achieve this enhancement, a change of approximately 0.3 V in the donor potential would be required upon binding of the analyte as indicated by Figure 3. Furthermore, a comparison with the experimental data of the cyano-substituted compound series listed in Table 3 and 4 shows that the fluorescence enhancement factor steadily increases with increasing ET driving force as also implied by Figure 3. Despite the fact that protonation of the aniline nitrogen is expected to yield a similarly large $\Delta E(D^+/D)$ for all compounds, the fluorescence enhancement factors vary over more than 2 orders of magnitude, underscoring the importance of tuning the ET driving force. To achieve the optimal fluorescence enhancement, the ET driving force must be adjusted within a narrow range centered around 0.55 ± 0.06 eV. In summary, the contour plot of Figure 3 outlines a generalized strategy for optimizing the fluorescence enhancement of triarylpyrazoline-based PET sensors. By systematically increasing the number of fluoro substituents attached to the 1-aryl ring, the ET driving force can be gradually increased until the maximum contrast enhancement is achieved. Driving forces beyond the optimal point should be avoided as they would lead to a reduced contrast ratio and quantum yield.

Quantum Chemical Calculations. The impetus for performing computational studies was 2-fold: the experimental data of the compound series **1** and **2** offered a valuable framework to gauge the ability of quantum chemical methods for predicting two of the ET key parameters, the acceptor potential $E(A/A^-)$ and the excited-state energy ΔE_{00} , by means of linear free energy relationships (LFERs). In addition, we hoped to gain insight into the unexpected reduction of the quantum yield upon protonation of derivatives **3**.

Reduction Potentials. The half-wave acceptor potential $E(A/A^-)$ for the one-electron reduction of the fluorophore is related to its gas-phase adiabatic electron affinity (AEA) according to the following relationship:

$$E(A/A^-) = \text{AEA} - \Delta\Delta G_{\text{sol}} - E_{\text{ref}} \quad (12)$$

where $\Delta\Delta G_{\text{sol}}$ is the energy difference between the neutral and reduced molecule in the gas phase and in solution and E_{ref} is the potential of the reference electrode.²² The implicit prediction of standard reduction potentials is in principle possible on the basis of ab initio molecular orbital theory or density functional theory to calculate AEA in combination with a solvation model that accounts for $\Delta\Delta G_{\text{sol}}$.²³ Because the latter is expected to vary within a narrow range for molecules with a similar degree of charge delocalization,²⁴ solution reduction potentials can be directly correlated with gas-phase electron affinities through a simple LFER.²⁵ Given the uniform molecular architecture of

the triarylpyrazoline compound library, we deemed that this approach should be well suited for correlating the experimental redox potentials with computational data. Using density functional theory, adiabatic gas-phase EAs were calculated as the difference between the total energies of the geometry-optimized anionic and charge-neutral fluorophores. We are aware that DFT typically yields positive anion HOMO energies, thereby implying instability toward electron detachment;²⁶ however, numerous detailed studies have concluded that, despite the presence of positive HOMO energies of the anion, DFT can indeed provide reasonable estimates of molecular EAs.²⁷ The electronic energies were corrected with the respective scaled harmonic ZPVEs.¹⁸ To evaluate the effects of the basis set size on the linear regression, the total energies were computed at the B3LYP/6-31+G(d) and B3LYP/6-311+G(2d,2p) levels of theory. In all cases, ZPVE corrections were obtained from frequency calculations of geometries optimized at the B3LYP/6-31+G(d) level. A list of the relevant computational results is given in Table 5, and selected regression analyses are shown in Figure 4.

As illustrated with Figure 4a, the adiabatic EAs computed with the larger basis set at the B3LYP/6-311+G(2d,2p) level correlated well with the experimental reduction potentials. Linear regression analysis yielded a mean unsigned error (MUE) of 0.02 eV and a good correlation coefficient of 0.995. Using the less expensive basis set 6-31+G(d), the EAs were estimated to be slightly smaller across all compounds; however, the linear regression yielded a similarly good fit with a correlation coefficient of 0.993 and an MUE of 0.03 eV. In both cases, however, the slope deviated considerably from unity, which may reflect either poor performance of DFT for modeling the AEAs or perhaps differences in solvation energies that are not captured by this approach. To further explore the latter possibility, we estimated the differential solvent stabilization energy $\Delta\Delta G$ on the basis of the semiempirical solvation model SM5.42R/AM1 developed by Cramer, Truhlar, and co-workers.²⁸ This model was parametrized to yield accurate solvation free energies at DFT gas-phase solute geometries. The values listed in Table 5 were thus computed from geometries optimized at the B3LYP/6-311+G(2d,2p) level of theory. The reduction potentials were calculated relative to the potential $E_{\text{ref}}(\text{Fc}^{+/0})$ of the ferrocenium cation in acetonitrile according to eq 12. To obtain $E_{\text{ref}}(\text{Fc}^{+/0})$ in acetonitrile, we selected a value of 4.52 V for the SHE as recommended by Cramer and Truhlar for a nonaqueous solution²⁹ and added 0.624 V for conversion to the $\text{Fc}^{+/0}$ reference,³⁰ which gives a reference potential of 5.14 V. The reduction potentials computed by this approach agreed well with the experimental values, yielding an MUE of 0.029 V, a good correlation coefficient of 0.992, and a slope that is closer to unity compared to the previous AEA gas-phase correlation. Inspection of the computed differential solvent stabilization energies reveals a steady decrease across both compound series by approximately 0.2 eV (Supporting Information), which is presumably due to the poorer solvation of the anion with an increasing degree of fluorine substitution.

In principle, the LUMO energies of the neutral species should also reflect differences in electron affinities, thus offering a simple approach for correlating the experimental reduction

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Table 5. Computed Gas-Phase LUMO Energies, Adiabatic Electron Affinities, Reduction Potentials, and Excited-State Energies for Pyrazoline Fluorophores **1a–j** and **2a–j**

compd	LUMO energy ^a (eV)			adiabatic EA ^a (eV)		reduction potential (V)		S ₁ energy (eV)	
	HF/6-311+G(2d,2p)	B3LYP/6-31+G(d)	B3LYP/6-311+G(2d,2p)	B3LYP/6-31+G(d)	B3LYP/6-311+G(2d,2p)	SM5.42R ^b B3LYP/6-311+G(2d,2p)	exptl ^c	TD-DFT B3LYP/6-311+G(2d,2p)	exptl ^d
1a	1.47	-1.47	-1.50	0.45	0.49	-2.79	-2.79	3.41	3.09
1b	1.54	-1.51	-1.61	0.51	0.55	-2.76	-2.78	3.50	3.19
1c	1.45	-1.59	-1.66	0.58	0.61	-2.78	-2.76	3.46	3.17
1d	1.42	-1.55	-1.63	0.53	0.57	-2.81	-2.77	3.43	3.09
1e	1.48	-1.60	-1.69	0.59	0.61	-2.80	-2.75	3.49	3.20
1f	1.49	-1.66	-1.74	0.65	0.67	-2.71	-2.70	3.52	3.23
1g	1.38	-1.71	-1.80	0.69	0.71	-2.75	-2.71	3.54	3.24
1h	1.43	-1.73	-1.80	0.72	0.74	-2.76	-2.69	3.49	3.22
1i	1.46	-1.64	-1.71	0.65	0.67	-2.83	-2.72	3.77	3.46
1j	1.39	-1.68	-1.74	0.73	0.70	–	-2.66	3.79	3.51
2a	1.02	-2.26	-2.34	1.19	1.23	-2.34	-2.26	3.11	2.79
2b	1.02	-2.28	-2.37	1.24	1.28	-2.34	-2.20	3.23	2.89
2c	0.91	-2.37	-2.45	1.30	1.34	-2.33	-2.21	3.15	2.86
2d	0.94	-2.33	-2.40	1.26	1.30	-2.36	-2.24	3.10	2.79
2e	0.94	-2.35	-2.42	1.31	1.34	-2.36	-2.21	3.23	2.91
2f	0.90	-2.41	-2.48	1.36	1.40	-2.26	-2.18	3.25	2.93
2g	0.82	-2.48	-2.56	1.41	1.44	-2.29	-2.16	3.21	2.93
2h	0.84	-2.46	-2.53	1.42	1.45	-2.29	-2.17	3.25	2.94
2i	0.95	-2.39	-2.48	1.34	1.37	-2.28	-2.21	3.49	3.15
2j	0.91	-2.43	-2.50	1.39	1.41	-2.34	-2.22	3.51	3.19
slope ^e	-0.977	-0.681	-0.677	0.723	0.717	1.15	–	1.022	–
intercept ^e	-1.309	-3.827	-3.870	-3.168	-3.182	0.469	–	-0.382	–
R ^e	0.987	0.988	0.993	0.993	0.995	0.992	–	0.995	–
MUE ^f	0.034	0.022	0.024	0.026	0.021	0.029	–	0.026	–

^a At the B3LYP/6-31+G(d) geometry. ^b According to eq 12. $\Delta\Delta G_{\text{sol}}$ calculated on the basis of the SM5.42R/AM1 solvent model.²⁸ Adiabatic EA at the B3LYP/6-311+G(2d,2p)//B3LYP/6-31+G(d) level of theory. ^c $E_{1/2}$ reduction potentials vs Fc/Fc⁺ in CH₃CN, 0.1 M Bu₄NPF₆, 298 K. ^d Excited-state energy ΔE_{00} (Table 1). ^e Regression analysis of computed vs experimental data. ^f Mean unsigned error.

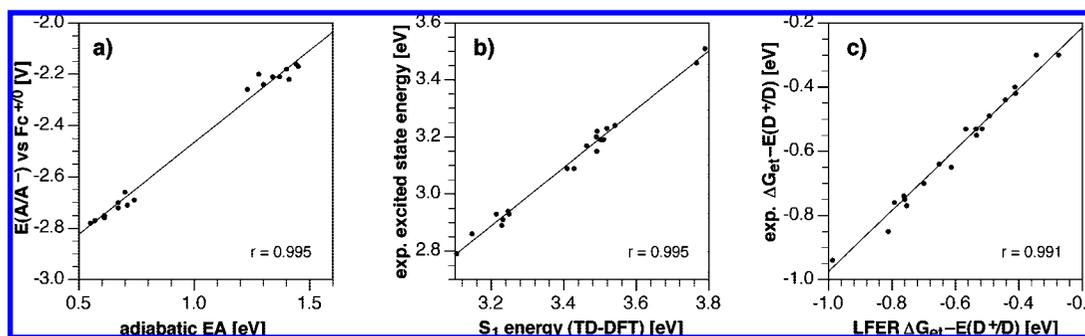


Figure 4. LFERs between selected computational and experimental data for compounds **1** and **2**. (a) Correlation of the gas-phase adiabatic electron affinity with experimental solution-phase reduction potentials. (b) Correlation of the vertical excitation energy of the lowest excited singlet state S₁ based on TD-DFT with the experimental excited-state energies. (c) Correlation between computed data according to the LFER of eq 13 and the experimental data. The computational data were calculated at the B3LYP/6-311+G(2d,2p)//B3LYP/6-31+G(d) level of theory. The slope, intercept, and MUE for all correlations are listed in Table 5.

potentials with computational data. Linear regression analysis indeed yielded surprisingly good correlations for both basis sets with MUEs at 0.024 and 0.026 V and a correlation coefficient of 0.993.

Excited-State Energies. The second key parameter that governs the photoinduced electron transfer thermodynamics is the excited-state energy ΔE_{00} , which corresponds to the energy difference between the geometry-optimized ground and lowest excited states corrected by the corresponding ZPVEs. While the computation of vertical excited-state energies is possible within 0.1–0.2 eV accuracy using time-dependent density functional theory (TD-DFT), the estimation of ΔE_{00} relies on excited-state geometries which are difficult to obtain with reliable accuracy. Given this limitation, we again applied a simple LFER for correlating the experimental ΔE_{00} energies with the TD-DFT vertical excitation energies on the basis of

the ground-state geometries. As shown in Figure 4b, linear regression analysis between the experimental and computational data yielded a slope close to unity with a good correlation coefficient of 0.989 and an MUE of 0.026 eV.

Finally, to estimate the electron transfer driving force for a given donor potential $E(D^+/D)$, the empirical LFER for the reduction potential and excited-state energy can also be combined in a single correlation, giving the empirical relationship

$$\Delta G_{\text{et}} (\text{eV}) = E(D^+/D) - 0.717(\text{AEA}) - 1.002S_1 + 3.526 \quad (13)$$

(29) Kelly, C. P.; Cramer, C. J.; Truhlar, D. G. *J. Phys. Chem. B* **2007**, *111*, 408–422.

where AEA is the adiabatic electron affinity computed at the B3LYP/6-311+G(2d,2p)//B3LYP/6-31+G(d) level and S_1 is the vertical excitation energy obtained from TD-DFT at the same level of theory.

A plot of the computed LFER data against the experimental values yielded an overall good correlation with $r = 0.991$ and $MUE = 0.023$ eV (Figure 4c). While we have not tested the accuracy of the correlation with compounds that were not used in the LFER, the computational data closely reflect the experimental trends over the entire set of 20 fluorophores. Hence, on the basis of the computed AEA and S_1 , it should be possible to prescreen pyrazoline fluorophores with various substituent combinations and ultimately identify those candidates whose electron transfer parameters would best match a given donor potential for an optimal contrast enhancement.

Oxidative Electron Transfer. As previously mentioned, compounds **3a**, **3b**, and **3d** do not exhibit an increase but rather a decrease in quantum yield upon acidification. Protonation mutates the aniline functionality into a strong σ -acceptor that might potentially trigger an oxidative rather than reductive electron transfer from the fluorophore. To explore this hypothesis, we performed gas-phase TD-DFT calculations of fluorophores **3a** and **4a** in the neutral and protonated state. Figure 5 illustrates the energy ordering of the three lowest excited states for each species. To visualize the nature of each state, electron detachment and attachment densities³¹ were plotted next to the corresponding energy levels. In the case of neutral fluorophore **3a**, an emissive state with charge transfer character resides at the lowest level followed by an ET state involving the aniline moiety as the electron donor (Figure 5a). In agreement with the measured quantum yield of 0.44 (Table 3), the energetically higher lying ET state is poorly accessible for nonradiative deactivation. For neutral fluorophore **4a** the energy ordering is reversed (Figure 5b). Consistent with the measured quantum yield of 0.014 (Table 3), the ET state lies below the charge transfer state, thus offering an energetically accessible nonradiative deactivation channel. Upon protonation of the aniline nitrogen, two states with ET character also now appear below the charge transfer state of fluorophore **3a** but not of **4a**. The detachment/

attachment densities indicate that both of these states involve an oxidative electron transfer from the fluorophore to the aniline moiety, which now acts as an electron acceptor rather than a donor. Whereas two states with analogous character are also observed for fluorophore **4a**, both of them reside at significantly higher energy compared to the lowest emissive charge transfer state and therefore are not expected to contribute significantly to nonradiative deactivation. Consistent with this model, fluorescence emission is switched on upon protonation of **4a**, but reduced in the case of **3a**. While the gas-phase calculations are not expected to quantitatively model the excited-state levels in solution, the results qualitatively support an oxidative electron transfer from the fluorophore π -system to the aniline cation upon protonation of **3a**.

Conclusions

Owing to their unusual electronic structure, 1,3,5-triarylpyrazolines offer the opportunity to differentially tune two of the key parameters that govern the PET thermodynamics, the excited-state energy ΔE_{00} and the acceptor potential $E(A/A^-)$. By varying the number and attachment positions of fluoro substituents, ΔE_{00} can be adjusted over a broad range without significantly altering the acceptor potential $E(A/A^-)$. On the basis of this strategy, the ET driving force can be predictably tuned to match the donor potential $E(D^+/D)$ of a given cation receptor and thus employed to optimize the contrast ratio of the sensor response. The experimentally measured D–A coupling and reorganization energy may be used to define an approximate potential map for finding the optimal ET driving force. Furthermore, semiempirical LFER correlations offer a useful tool to estimate the acceptor potential, excited-state energy, and ET driving force with a mean unsigned error of 0.03 eV and thus to computationally prescreen substituent patterns for their suitability prior to actual synthesis of the corresponding fluorophores. The current LFER training set has been calibrated in acetonitrile and is limited in size; however, it could certainly be expanded to different solvent systems including a broader substituent coverage. The differential

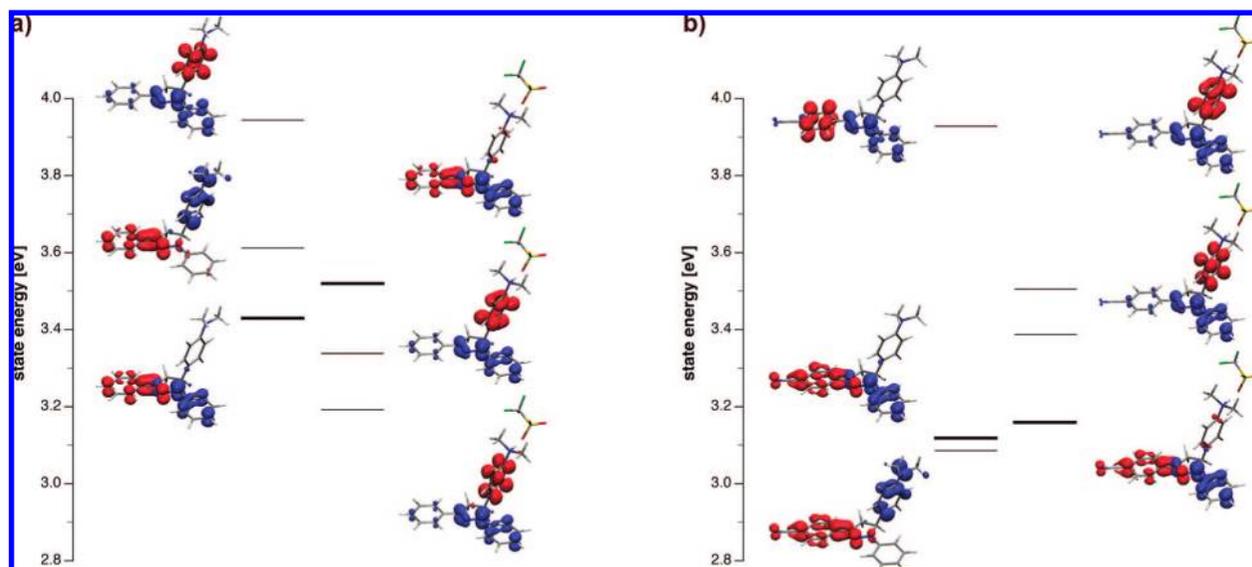


Figure 5. Gas-phase excited-state manifold of pyrazoline fluorophore **3a** (a) and **4a** (b) under neutral conditions (left) and upon protonation with trifluoroacetic acid (right) (TD-DFT calculations at the B3LYP/6-311+G(2d,2p)//B3LYP(6-31+(d) level of theory). Detachment (blue) and attachment (red) density plots are shown next to the energy level of each state. States with significant oscillator strength are marked with bold lines.

tuning ability of the ET parameters in triarylpyrazoline fluorophores is a direct consequence of the spatial separation of the HOMO and LUMO densities while still providing sufficient overlap and therefore a substantial absorption cross section. It is conceivable that the outlined rational design strategy could be applied to other fluorophore platforms that exhibit similar spatial separations of the HOMO and LUMO densities.³²

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Supporting Information Available: Synthesis and characterization of derivatives **1–4**, complete ref 15, and additional information as mentioned in the text. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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